Differences in Hydration and Association of Helical Boc-Val-Ala-Leu-Aib-Val-Ala-Leu-(Val-Ala-Leu-Aib)₂-OMe·xH₂O in Two Crystalline Polymorphs[†]

Isabella L. Karle,*,[‡] Judith L. Flippen-Anderson,[‡] Muppalla Sukumar, and Padmanabhan Balaram

Laboratory for the Structure of Matter, Naval Research Laboratory, Washington, D.C. 20375-5320, and Molecular Biophysics Unit, Indian Institute of Science, Bangalore 560 012, India

Received March 30, 1992

The 15-residue apolar peptide, Boc-Val-Ala-Leu-Aib-Val-Ala-Leu-(Val-Ala-Leu-Aib)₂-OMe has been crystallized from 2-propanol-water (form I). The crystal parameters for I are as follows: $C_{74}H_{133}N_{15}O_{18}\cdot 2H_2O$, space group P2, a = 9.185 (6) Å, b = 47.410 (3) Å, c = 10.325 (9) Å, β = 91.47 (2)°, Z = 2, R = 6.3% for 4532 reflections observed $> 3\sigma(F)$, resolution 0.94 Å. The structure is almost completely α -helical with eleven 5-1 hydrogen bonds and one 4-1 hydrogen bond near the N-terminus. The structure has been compared with a polymorph (form II) obtained from methanol-water (Karle, I. L.; Flippen-Anderson, J. L.; Uma, K.; Sukumar, M.; Balaram, P., J. Am. Chem. Soc. 1990, 112, 9350–9356). The two forms differ in the extent of hydration; form I contains two water molecules in the head-to-tail region of helical columns, while form II is more extensively solvated, with the equivalent of 7.5 water molecules. The three-dimensional packing of helices is completely parallel in I and antiparallel in II.

 α -Aminoisobutyric acid (Aib) is an important constituent of many membrane active fungal peptides.^{1,2} Extensive studies of the stereochemistry of Aib-containing peptides³⁻⁶ have been motivated by attempts to develop structure-activity correlations for membrane channel forming peptides^{7,8} and the possibility of using stereochemically constrained residues in designing conformationally rigid analogs of biologically active peptides.⁹⁻¹² The ability of α -aminoisobutyryl residues to stabilize helical conformations in oligopeptides has permitted the detailed structural analysis of peptide helices in crystals

(5) Bosch, R.; Jung, G.; Schmitt, H.; Winter, W. Crystal Structure of $the \,\alpha\text{-}Helical \,Undecape ptide \,Boc\text{-}L\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aib\text{-}Aib\text{-}Ala\text{-}Glu(OBzl)\text{-}Ala\text{-}Aibb\text{-}Aibb\text{-}Aibb\text{-}Aibb\text{-}Aibb\text{-}Aibb\text{-}Aibb\text{-}Aibb\text{-}Aibb\text{-}Aibb\text{-}Ai$ Aib-Ala-Aib-Ala-OMe. Biopolymers 1985, 24, 961-978.

(6) Marshall, G. R.; Hodgkin, E. E.; Langs, D. A.; Smith, G. D.; Zabrocki, J.; Leplawy, M. T. Factors governing helical preference of peptides containing multiple α, α -dialkyl amino acids. *Proc. Natl. Acad. Sci. U.S.A.* 1990. 87. 487-491.

(8) Karle, I. L.; Flippen-Anderson, J. L.; Agarwalla, S.; Balaram, P. Crystal structure of [Leu1]zervamicin, a membrane ion-channel peptide: Implications for gating mechanisms. Proc. Natl. Acad. Sci. U.S.A. 1991, 88, 5307-5311

(9) Marshall, G. R.; Bosshard, H. E. Angiotensin II, Studies on the Biologically Active Conformation. Circ. Res. 1972, 30/31 (Suppl. II), 143-150.

(10) Jorgensen, E. C.; Rapaka, S. R.; Windridge, G. C.; Lee, T. C. Angiotensin II analogs. Stereochemical factors in the 5 position influencing pressor activity. J. Med. Chem. 1971, 14, 904-6.

(11) Sudha, T. S.; Balaram, P. Stabilization of β -turn conformations in enkephalins. a-Aminoisobutyric acid analogs. Int. J. Pept. Protein Res. 1983, 21, 381-388.

(12) Sukumar, M.; Raj, P. A.; Balaram, P.; Becker, E. L. Highly Active Chemotactic Peptide Analog Incorporating the Unusual Residue 1-Aminocyclohexanecarboxylic Acid at Position 2. Biochem. Biophys. Res. Commun. 1985, 128, 339-344.

at high resolution.^{13,14} These studies have provided several useful insights into packing and solvation of helices in the solid state, in addition to allowing the detailed stereochemical characterization (α , 3_{10} , or mixed $3_{10}/\alpha$) of helices.¹³⁻¹⁷ The stereochemical rigidity of Aib-containing peptide helices extends into solution,¹⁸ suggesting that these residues could be effectively employed in the construction of conformationally rigid, secondary structure modules, in the de novo design of synthetic protein mimics.¹⁹⁻²² This strategy to construct synthetic proteins rests on the ability to synthesize and characterize relatively long helical segments, which can then be assembled into larger structures.²³ This paper describes the structure of a model 15-residue peptide, Boc-Val-Ala-Leu-Aib-Val-Ala-Leu-(Val-Ala-Leu-Aib)2-OMe, crystallized from 2-pro-

(13) Karle, I. L.; Balaram, P. Structural Characteristics of α -Helical Peptide Molecules Containing Aib Residues. Biochemistry 1990, 29, 6747-6756.

(14) Toniolo, C.; Benedetti, E. The polypeptide 3₁₀-helix. Trends Biochem. Sci. (Pers. Ed.) 1991, 16, 350-353.

(15) Karle, I. L.; Flippen-Anderson, J. L.; Uma, K.; Balaram, H.; Balaram, P. α -Helix and mixed $3_{10}/\alpha$ -helix in cocrystallized conformers of Boc-Aib-Val-Aib-Aib-Val-Val-Aib-Val-Aib-OMe. Proc. Natl. Acad. Sci. U.S.A. 1989, 86, 765-769.

(16) Karle, I. L.; Flippen-Anderson, J. L.; Uma, K.; Balaram, P. Apolar Peptide Models for Conformational Heterogeneity, Hydration and Packing of Polypeptide Helices. Crystal Structure of Hepta- and Octapeptides Containing α-Aminoisobutyric Acid. Proteins: Struct. Funct.

Genet. 1990, 7, 62-73. (17) Pavone, C.; Toniolo, C.; Crisma, M. The Longest Regular Polypeptide 310 Helix at Atomic Resolution. J. Mol. Biol. 1990, 214, 633-635.

(18) Uma, K. Modular Design of Synthetic Protein Mimics. Con-struction of Helices. 1990, Ph.D. Thesis, Indian Institute of Science,

Bangalore, India. Thesis Abstr. J. Ind. Inst. Sci. 1991, 71, 395–398.
(19) Balaram, P. Peptides as Bioorganic Models. Proc. Ind. Acad. Sci.

Chem. Sci. 1984, 93, 703-717. (20) Karle, I. L.; Flippen-Anderson, J. L.; Uma, K.; Balaram, P. Modular Design of Synthetic Protein Mimics. Characterization of the Helical Conformation of a 13-Residue Peptide in Crystals. Biochemistry 1989, 28, 6696-6701.

(21) Karle, I. L.; Flippen-Anderson, J. L.; Uma, K.; Balaram, P. Helix Construction Using α -Aminoisobutyric Residues in a Modular Approach to Synthetic Protein Design. Curr. Sci. 1990, 59, 875-885. (22) DeGrado, W. F.; Raleigh, D. P.; Handel, T. De novo protein

design: what are we learning? Curr. Opinion Struct. Biol. 1991, 1, 984-993.

(23) Karle, I. L.; Flippen-Anderson, J. L.; Sukumar, M.; Uma, K.; Balaram, P. Modular Design of Synthetic Protein Mimics. Crystal Structure of Two Seven-Residue Helical Peptide Segments Linked by e-Aminocaproic Acid. J. Am. Chem. Soc. 1991, 113, 3952-3956.

[†] This paper is dedicated to Professor Ralph Hirschmann on the occasion of his 70th birthday. It has been a pleasure to know him. [‡] Naval Research Laboratory.

⁽¹⁾ Mathew, M. K.; Balaram, P. Alamethicin and related channel forming polypeptides. *Mol. Cell. Biochem.* 1983, 50, 47-64.

⁽²⁾ Krishna, K.; Sukumar, M.; Balaram, P. Structural chemistry and

membrane modifying activity of the fungal polypeptides zervamicins, antiamoebins and efrapeptins. *Pure Appl. Chem.* 1990, 62, 1417–1420. (3) Prasad, B. V. V.; Balaram, P. The Stereochemistry of Peptides Containing α -Aminoisobutyric Acid. CRC Crit. Rev. Biochem. 1984, 16, 307-347.

⁽⁴⁾ Toniolo, C.; Benedetti, E. Old and new structures from studies of synthetic peptides rich in C alpha, alpha-disubstituted glycines. ISI Atlas Sci.: Biochem. 1988, 1, 225-230.

⁽⁷⁾ Nagaraj, R.; Balaram, P. Alamethicin, a transmembrane channel. Acc. Chem. Res. 1981, 14, 356-362.

Table I. Diffraction Data for Two Polymorphs of Boc-Val-Ala-Leu-Aib-Val-Ala-Leu-(Val-Ala-Leu-Aib)₂-OMe-xH₂O

	I (present paper)	II (ref 24)
empirical formula	C74H133N15O18-2H2O	C74H133N15O18-7.5H2O
crystallizing solvent	2-propanol-H ₂ O	methanol-H ₂ O
crystal size, mm	$0.15 \times 0.25 \times 0.25$	$0.65 \times 0.30 \times 0.15$
space group	P2 ₁	$C222_{1}$
csll parameters		
a, Å	9.185 (6)	18.348 (5)
b, A	47.410 (3)	47.382 (11)
c. Å	10.325 (9)	24.157 (5)
β. deg	$\beta = 91.47(2)$	
vol. Å ³	4495	21001 (9)
Z	2	8
mol wt	1520.98 + 36.03	1520.98 + 135.12
density, g/cm	1.150	1.048
resolution, A	0.94	1.00
independent reflections	5724	5909
obsd refl $(F_n > 3\sigma(F))$	4532	3147
no. parameters refined	979	1045
no. data to no. parameters ratio	4.6:1.0	3.0:1.0
final R indices (obsd data), $\%$	6.3	10.6
goodness-of-fit	1.2	12.2
max diff, e/Å ³	0.35	0.36
min diff (hole), $e/Å^3$	-0.41	-0.31

panol-water (I) and compares the conformation and solvation with that of a polymorph (II) obtained earlier from methanol-water.²⁴ While the crystals of polymorph I are stable for several weeks in the dry state, the crystals of polymorph II exist as stable colorless plates only when surrounded by a large amount of mother liquor. The two polymorphs differ in the extent of hydration and also show subtle differences in the peptide helix with respect to the intrahelical hydrogen bonding pattern near the N-terminus.

Experimental Section

The model peptide was synthesized by a conventional solution-phase procedure by a fragment condensation approach.²⁵ After purifying the peptide by high-performance liquid chromatography, crystals were grown by slow evaporation from a 2-propanol- H_2O solution.

X-ray diffraction data were collected with Cu K α radiation from a dry crystal at 228 K on an automated four-circle diffractometer with a graphite monochromator (Nicolet R3). Three reflections used as standards, monitored after every 97 measurements, remained constant within 4% during the data collection. The θ -2 θ scan technique was used with $2\theta_{max} = 110^\circ$, a scan width of 2.0° $+ 2\theta(\alpha_2) - 2\theta(\alpha_1)$, scan speed of 14.6°/min and index ranges h: -9 to +10; k: 0 to 49; and l: -10 to +1. Cell parameters for the two polymorphs and other diffraction data are shown in Table I. The present structure was solved by using the molecule in the earlier structure as a vector search model in the PATSEE computer programs.²⁷ Due to the large number of parameters, full-matrix anisotropic least-

(25) Balaram, H.; Sukumar, M.; Balaram, P. Stereochemistry of α -Aminoisobutyric Acid Peptides in Solution: Conformations of Decapeptides with a Central Triplet of Contiguous L-Amino Acids. Biopolymers 1986, 25, 2209–2223. (26) Egert, E.; Sheldrick, G. M. Search for a Fragment of Known

(26) Egert, E.; Sheldrick, G. M. Search for a Fragment of Known Geometry by Integrated Patterson and Direct Methods. Acta Crystallogr. Sect. A: Found. Crystallogr. 1985, 41, 262-268. squares refinement was performed on the C, N, and O atoms in two blocks (residues 1-7 and residues 8-15 with the two water oxygens refined in each block). The weighting scheme was $w = [\sigma^2(F) + 0.0010F^2]^{-1}$. Hydrogen atoms were placed in idealized positions, with N-H and C-H = 0.96 Å and allowed to ride with the C or N atom to which each was bonded for the final cycles of refinement. The thermal factor for the hydrogen atoms was fixed at $U_{\rm iso} = 0.125$. The side chain in Leu 14 has high thermal parameters and, in addition, the C³ atoms occupy all three conformational positions on the $C^{\gamma}(14)$ atom. Occupancies for atoms C⁵²(14) and C^{52'}(14) are $\sim^2/_3$ and $\sim^1/_3$ for sites with χ^2 near 50° and -72°, respectively. The final R factor is 6.3% for 4532 independent data measured > $3\sigma(F)$. Fractional coordinates are listed in the supplementary material, and torsional angles are listed in Table II.

Results

The Helix. The molecule folds into an almost complete α -helix with eleven 5 \rightarrow 1 hydrogen bonds in the crystal form I from 2-propanol- H_2O . The 5-1 hydrogen bonds run successively beginning with N(5)H; N(4)H does not participate in any hydrogen bonding and N(3)H makes a $4 \rightarrow 1$ hydrogen bond with carbonyl O(0) from the Boc group (Table III). The initial 3_{10} turn stabilized by a $4 \rightarrow 1$ hydrogen bond at the N-terminus of a long helix is a fairly common feature in peptides and proteins.²⁸ The φ and ψ values (Table II) lie close to the values expected for ideal α -helices with somewhat greater distortions for the Val-(12), Ala(13) and Leu(14) residues. The crystal form II reported earlier²⁴ adopts a complete α -helical conformation with twelve $5 \rightarrow 1$ hydrogen bonds. The φ and ψ values for form II are also listed in Table II for comparison. The overall conformation in the two forms is very similar with the four Leu side chains extended on one side of the helix and the four Val side chains extended on the other side. Only the terminal carbomethoxy group differs by a 180° rotation in the two crystalline forms.

Head-to-Tail Hydrogen Bonding. Figure 1 shows a comparison of the head-to-tail hydrogen bonding in crystals I and II.²⁴ The peptides are shown in nearly the same orientation. The direct N(1)H-O(13) hydrogen bond is the same in both. Water molecule W(2) is in a similar location in both, acting as a hydrogen bonding intermediary between N(2)H and O(13). Water molecules W(1) in I and W(5) in II both form hydrogen bonds with O(12), but their other hydrogen bonds are quite different. There are only two cocrystallized water molecules in I but in II there are six water molecules in the head-to-tail region, three of which are disordered among six water sites. (Only one position of each disordered water for II is shown in Figure 1.) As mentioned in the above section, a complete α -helix is formed in II.²⁴ In I the helix appears to adjust to fewer waters for hydrogen bond mediation by switching to a 3_{10} -type for N(3)H and by N(4)H not participating in any hydrogen bonding.

Interhelical Side-Chain Contacts. Figure 2 shows Leu.--Val nearest contacts in a sheet in crystal I where the molecules are related by translation. Figure 3 shows molecules in a sheet in crystal II, where the molecules are related by a 2-fold rotation between Val.---Val contacts or between Leu.--Leu contacts. The right molecule in each figure is in nearly the same orientation. The dotted lines

⁽²⁴⁾ Karle, I. L.; Flippen-Anderson, J. L.; Uma, K.; Sukumar, M.; Balaram, P. Modular Design of Synthetic Protein Mimics. Crystal Structures, Assembly, and Hydration of Two 15- and 16-Residue Apolar, Leucyl-Rich Helical Peptides. J. Am. Chem. Soc. 1990, 112, 9350–9356.

⁽²⁷⁾ Sheldrick, G.M. SHELXTL PLUS Version 4.1. An Integrated System for Solving, Refining and Displaying Crystal Structures from Diffraction Data (1990), Siemens Analytical X-Ray Instruments, Madison, WI.

⁽²⁸⁾ Baker, E. N.; Hubbard, R. E. Hydrogen Bonding in Globular Proteins. Prog. Biophys. Mol. Biol. 1984, 44, 97-179.

Table II. Torsional Angles^{a.b}

	φ	Ý	ω	x ¹	x ²
Val(1)	-51.8 (-55.5)°	-45.6 (-60.2)	-173.2	-67.4, 169.2	
Ala(2)	-63.6 (-65.5)	-32.5 (-38.5)	179.3		
Leu(3)	-65.9 (-64.8)	-51.7 (-40.3)	-179.3	-179.6	-173.2, 65.5
Aib(4)	-55.2 (-51.9)	-48.4 (-50.0)	-176.9		·
Val(5)	-63.0 (-61.8)	-50.1 (-45.8)	-176.3	-67.5, 168.7	
Ala(6)	-61.1 (-60.3)	-42.9 (-39.5)	-177.3		
Leu(7)	-68.6 (-67.7)	-38.9 (-34.0)	175.8	- 64 .0	-178.9, -55.1
Val(8)	-63.9 (-69.5)	-45.1 (-45.1)	-179.2	-67.2, 169.9	
Ala(9)	-60.4 (-58.9)	-45.0 (-42.7)	178.8		
Leu(10)	-61.9 (-62.4)	-52.1 (-46.8)	-173.3	-73.5	162.8, -71.3
Aib(11)	-57.2 (-51.6)	-49.5 (-46.0)	-167.7		
Val(12)	-79.5 (-72.5)	-31.8 (-30.6)	178.3	-179.9, -60.5	
Ala(13)	-75.3 (-67.8)	-42.0 (-34.4)	-172.7		
Leu(14)	-81.2 (-87.3)	-28.5 (-20.0)	-169.3	-82.9	170.4, 49.5, 71.8/
Aib(15)	-56.3 (-57.5)	$-41.0 (148.2)^d$	-171.5 ^e		· · ·

^a The torsion angles for rotation about bonds of the peptide backbone (ϕ , ψ , and ω) and about bonds of the amino acid side chains (χ^1 and χ^2) are described in ref 38. ^b ESD's are ~1.0°. Values in parentheses correspond to form II crystallized from CH₃OH-H₂O.²⁴ ^c C'(0), N(1), C^a(1), C'(15), C'(15), O(OMe). ^e C^a(15), C'(15), O(OMe). ^f Disorder among C⁵(14) atom positions.

Table III. Hydrogen Bonds

type	donor	acceptor	N…O, Å	HO,ª Å	C==ON angle, deg
heat-to-tail	N(1)	O(13) ^b	3.029	2.16	165.7
peptide-to-water	N(2)	W(2)°	2.819	1.93	
4-→1	N(3)	O(0)	3.054	2.37	125.5
	$N(4)^d$				
5-→1	N(5)	O(1)	3.036	2.15	164.1
	N(6)	O(2)	2.941	2.08	146.8
	N(7)	O(3)	2.932	2.06	159.2
	N(8)	O(4)	3.151	2.27	152.4
	N(9)	O(5)	2.986	1.88	157.1
	N(10)	O(6)	2.933	2.06	154.9
	N(11)	O(7)	2.911	2.04	149.8
	N(12)	O(8)	3.001	2.13	156.8
	N(13)	O(9)	3.031	2.20	148.6
	N(14)	O(10)	2.976	2.11	156.4
	N(15)	0(11)	3.156	2.40	144.0
			00,		
			Ā		
water-to-peptide	W (1)	O(12) ^e	2.912		
or water	W(1)	O(14) ^f	2.871		
	W(2)	W(1)	2.749		
	W(2)	O(13) ⁶	2.960		

^a The H atoms were placed in idealized positions with the N-H distance equal to 0.96 Å. ^b Symmetry equivalent 2 - x, $\frac{1}{2} + y$, 1 - z to coordinates listed in the supplementary material. ^c Symmetry equivalent 1 + x, y, z to coordinates listed in the supplementary material. ^d Atoms N(4) and O(15) do not participate in hydrogen bonding; N(4)--O(0) = 3.42 Å, angle C-O--N = 162°. ^c Symmetry equivalent 1 - x, $\frac{1}{2} + y$, 1 - z to coordinates listed in the supplementary material. ^f Symmetry equivalent 1 - x, $\frac{1}{2} + y$, 2 - z to coordinates listed in the supplementary material.

show intermolecular C···C contacts (~3.8 to 4.1 Å, a very few are <3.8 Å). In crystal I there are eleven such contacts while in crystal II, seven such contacts occur between Val···Val, or if the Leu···Leu contacts on the other side of the helix are examined, there are eight. In either case there are fewer C···C close approaches in II than in I. Even taking into account the occurrence of water molecules in some of the cavities (where C···O approaches are ≥ 3.9 Å), the packing is looser in II than in I. The larger calculated



Figure 1. Head-to-tail hydrogen bonds between two molecules (related by a 2-fold screw axis) of Boc-Val-Ala-Leu-Aib-Val-Ala-Leu-(Val-Ala-Leu-Aib)₂-OMe in polymorph I (P_{1} , with $2H_{2}O$) (present paper) and polymorph II ($C222_{1}$, with $7.5H_{2}O$) (see ref 24). The large black dots represent water molecules and the dashed lines represent hydrogen bonds.

density, 1.15 g cm⁻³ in I as compared to 1.05 g cm⁻³ in II is consistent with more efficient packing in I.

Helix Assembly in Crystals. Peptide helices possess appreciable macrodipole moments.^{29–31} Antiparallel helix packing in proteins has been rationalized by invoking

⁽²⁹⁾ Wada, A. The alpha-helix as an electric macro-dipole. Adv. Biophys. 1976, 9, 1-63. (30) Hol, W. G. J.; Halie, L. M.; Sander, C. Dipoles of the α -helix and

⁽³⁰⁾ Hol, W. G. J.; Halie, L. M.; Sander, C. Dipoles of the α -helix and β -sheet: Their role in protein folding. Nature (London) 1981, 294, 532–536.

⁽³¹⁾ Schwarz, G.; Savko, P. Structural and dipolar properties of the voltage-dependent pore former alamethicin in octanol/dioxane. *Biophys. J.* 1982, *39*, 211.



Figure 2. Two molecules of the 15-residue peptide in polymorph I that are related by translation along the ac diagonal. Dotted lines indicate nearest C…C distances (\sim 3.8 to 4.1 Å) between neighboring molecules. Mostly Leu-Val van der Waals contacts are involved.

dipole-dipole interactions as an important determinant in protein folding.³⁰ Electrostatic energy calculations for helical peptides packed in crystals have led to the conclusion that antiparallel helix packing is overwhelmingly favored.³² However, this has not been the case for 10-16 residue apolar peptide helices where many instances of completely parallel packing of helices has been observed in crystals. In a number of cases, the same peptide crystallized in different crystal forms has been found to have all parallel assemblies of helices in one crystal form and an antiparallel assembly in a different crystal form.^{33,34} Similarly, for the present 15-residue peptide, there is completely parallel packing of helices in crystal I. In crystal II,²⁴ although there is parallel packing, in the sense of the helix direction, that occurs in sheets of one molecule thickness as shown in Figure 3, the adjacent sheets assemble in an antiparallel motif. The existence of a dipole moment in a helix does not appear to influence the packing direction of helices.

The demonstration that peptide helices can assemble in a completely parallel fashion³⁵ was instrumental in



Figure 3. Two molecules of the 15-residue peptide in polymorph II²⁴ related by a 2-fold rotation axis. Dotted lines indicate nearest C…C distances (\sim 3.8 to 4.1 Å) between neighboring molecules. Val---Val and Val---Aib van der Waals contacts are involved.

designing a peptide having chymotrypsin-like esterase activity.^{36,37} In this peptide, four short parallel helical peptides were linked covalently at their carboxyl ends while their amino ends bore the serine protease catalytic site residues serine, histidine, and aspartic acid.

Concluding Remarks

A 15-residue apolar peptide has been shown to crystallize in different crystal forms in which completely parallel packing of helices occurs in one, and antiparallel packing occurs in the other. The association of helices is not governed by particular selectivities of the various side chains. Although both polymorphs contain water of crystallization, there is much more water around the headto-tail region (and some additional water molecules laterally between the helices) in the crystal in which the packing is less efficient. Nevertheless, the conformation of the peptide molecule is nearly the same in both, that is, completely α -helical in one and one $5 \rightarrow 1$ hydrogen bond replaced by a $4 \rightarrow 1$ hydrogen bond in the other. It is important to note that the choice of solvent, and conse-

⁽³²⁾ Hol, W. G. J.; de Maeyer, M. C. H. Electrostatic interactions between α -helix dipoles in crystals of an uncharged helical undecapeptide. Biopolymers 1984, 23, 809-817.

⁽³³⁾ Karle, I. L.; Flippen-Anderson, J. L.; Uma, K.; Balaram, P. Parallel and Antiparallel Aggregation of α -Helices. Crystal Structures of Two Apolar Decapeptides X-Trp-Ile-Ala-Aib-Ile-Val-Aib-Leu-Aib-Pro-OMe (X = Boc, Ac). Int. J. Pept. Protein Res. 1990, 35, 518-526.

⁽³⁴⁾ Karle, I. L.; Flippen-Anderson, J. L.; Uma, K.; Balaram, P. Helix Aggregation in Peptide Crystals: Occurrence of Either All Parallel or Antiparallel Packing Motifs for α -Helices in Polymorphs of Boc-Aib-Ala-Leu-Ala-Leu-Aib-Leu-Ala-Leu-Aib-OMe. Biopolymers 1990, 29, 1835-1845.

⁽³⁵⁾ Karle, I. L.; Sukumar, M.; Balaram, P. Parallel packing of a-Helices in crystals of the zervamicin IIA analog Boc-Trp-Ile-Ala-Aib-Ile-Val-Aib-Leu-Aib-Pro-OMe-2H2O. Proc. Natl. Acad Sci. U.S.A. 1986, 83, 9284-9288

 ⁽³⁶⁾ Stewart, John M., private communication.
(37) Hahn, K. W.; Klis, W. A.; Stewart, J. M. Design and Synthesis of a peptide having chymotrypsin-like esterase activity. Science 1990, 248, 1544-1547

⁽³⁸⁾ IUPAC-IUB Commission on Biochemical Nomenclature Abbreviations and Symbols for the Description of the Conformation of Polypeptide Chains. Biochemistry 1970, 9, 3471-3479.

Crystalline Peptide Helices

quently the amount of cocrystallized water, has a profound effect on the stability of the crystals.

Acknowledgment. This research was supported by the Office of Naval Research, National Institutes of Health Grant GM30902 and in part by a grant from the Department of Science and Technology, India. K.U. was supported by a fellowship from the Council of Scientific and Industrial Research, India.

Journal of Medicinal Chemistry, 1992, Vol. 35, No. 21 3889

Supplementary Material Available: Tables of atomic coordinates, bond lengths, bond angles, anisotropic displacement coefficients, and H atom coordinates (10 pages); observed and calculated structure factors (21 pages). Ordering information is given on any current masthead page.

Registry No. I, 130378-94-8; I·2H₂O, 143294-07-9.